

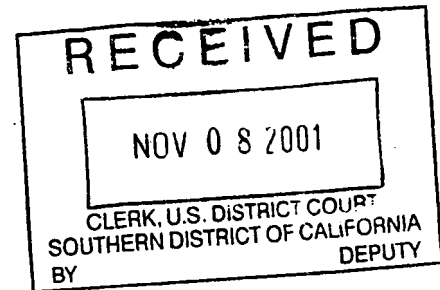
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14 UNITED STATES DISTRICT COURT  
15 SOUTHERN DISTRICT OF CALIFORNIA

17 GEN-PROBE, INCORPORATED,  
18 Plaintiff,  
19 v.  
20 VYSIS, INC.,  
21 Defendant.

CASE NO. 99CV 2668H (AJB)

**VYSIS' SUPPLEMENTAL  
STATEMENT OF DISPUTED FACTS  
IN OPPOSITION TO GEN-PROBE'S  
MOTION FOR PARTIAL SUMMARY  
JUDGMENT OF  
NONINFRINGEMENT UNDER THE  
DOCTRINE OF EQUIVALENTS**

Date: November 19, 2001  
Time: 10:30 a.m.  
Place: Courtroom 1

22 Defendant Vysis, Inc. respectfully submits the following supplemental statement of disputed  
26 material facts, together with supporting evidence, in support of its Opposition to Gen-Probe's  
27 Motion for Partial Summary Judgment of Noninfringement Under the Doctrine of Equivalents.  
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<b>GEN-PROBE ALLEGED UNDISPUTED FACTS</b>	<b>DISPUTED FACTS AND SUPPORTING EVIDENCE</b>
1. Vysis has previously admitted that TMA is a sequence-specific amplification method and does not use methods of non-specific amplification.	Vysis did not dispute this assertion in its opposition to Gen-Probe's April 30, 2001 Motion for Partial Summary Judgment.
2. All of the claims of the '338 patent incorporate an "amplification" element. The Court's June 20th Order confirms that each of those claims and incorporated amplification elements literally encompasses only non-specific amplification techniques.	The Court's construction of the claims of the '338 patent is a legal question, not a factual one. Vysis contends that the Court's resolution of that question of law is legally incorrect.
3. The differences between specific amplification methods and non-specific amplification methods are substantial.	Disputed. See Persing Decl., ¶¶ 5-16.
4. The methods do not perform the same function in the same way to achieve the same result.	Disputed. See Persing Decl., ¶¶ 5-16.
5. Gen-Probe's TMA method functions to exponentially increase both the <b>absolute</b> and <b>relative</b> amount of a particular nucleic acid sequence of interest in a mixture of nucleic	No dispute.

1	acids.	
2		
3	6. In direct contrast, non-specific	In the context of the claims of the '338 patent,
4	amplification functions only to increase the	the amplification step increases both the
5	absolute amount of all nucleic acids present in	absolute and relative amount of the target
6	a sample and does not increase the relative	nucleic acid present in the tested sample. See
7	amount of a particular nucleic acid sequence	'338 patent; Mullis Dep. at 117.
8	of interest.	
9		
10	7. Vysis' own expert has admitted the	Vysis' expert has not opined that there is no
11	differences in function between specific	difference between specific and nonspecific
12	amplification and non-specific amplification.	amplification techniques, but has the opinion
13		that the differences are insubstantial. See
14	[N]on-specific amplification	Persing Decl. ¶¶ 5-16.
15	techniques amplify all of the nucleic	
16	acid in a sample, both target and	
17	non-target nucleic acid. Specific	
18	amplification techniques, <i>in</i>	
19	<i>contrast</i> , are intended to amplify	
20	only the target nucleic acid.	
21	8. When a particular nucleic acid sequence of	No dispute.
22	interest is contained in a mixture of nucleic	
23	acids in a clinical sample, TMA enables a	
24	person skilled in the art to exponentially copy	
25	the sequence of interest.	
26		
27	9. This makes it easy to determine whether or	No dispute.
28	not a pathogenic microorganism is hiding	

<p>1 among millions of other organisms in a 2 patient sample. 3</p>	
<p>4 10. Specific amplification is useful for 5 diagnostic purposes even without a target 6 capture step. In contrast, non-specific 7 amplification is <i>not</i> a viable diagnostic 8 method because it does not increase the 9 amount of a target nucleic acid relative to 10 everything else. Vysis' own expert witness 11 has admitted this important distinction: 12 13 14 Without the use of target capture 15 prior to amplification, <i>non-specific</i> 16 <i>amplification would not be a viable</i> 17 <i>technique for detecting target</i> 18 <i>nucleic acids in a sample</i> because, 19 as pointed out in the quoted 20 paragraph, non-specific 21 amplification causes the replication 22 of virtually any nucleic acid 23 sequence, including other irrelevant 24 nucleic acids in the sample.</p>	<p>Vysis disputes that non-specific amplification is "not a viable diagnostic method." Non- specific amplification is a viable diagnostic method when used in the context of claims of the '338 patent. May 25, 2001 Persing Decl., ¶ 11.</p>
<p>22 11. Therefore, Dr. Persing has admitted that 23 "without the invention [i.e., the combination 24 of a preliminary "target capture" step with 25 amplification], <i>only specific amplification</i> 26 <i>could be used.</i>" 27 28</p>	<p>Vysis disputes that the quoted section of Dr. Persing's May 25, 2001 Declaration was based on the assertions in Gen-Probe's Undisputed Fact No. 10.</p>

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<p>12. The enzymes and primers used in any amplification process can be specific or non-specific.</p>	<p>No dispute.</p>
<p>13. The primers used in Gen-Probe's specific TMA amplification method have been carefully selected by Gen-Probe's scientists and are generally designed to bind to specific, unique sequences in a DNA or RNA molecule.</p>	<p>No dispute.</p>
<p>14. In amplification processes, sequence-specific primers and enzymes such as those used in TMA play a role substantially different from non-specific primers and enzymes.</p>	<p>Disputed. See Persing Decl., ¶¶ 10-16.</p>
<p>15. This fact is well known to those of ordinary skill in the art.</p>	<p>Disputed. See Persing Decl., ¶¶ 10-16.</p>
<p>16. For example, specific primers and enzymes can function together to amplify a target nucleic acid only if the specific sequence of interest bound by the primer and/or recognized by the enzymes is present</p>	<p>Disputed. All nucleic acid amplification techniques have some degree of nonspecificity. See Persing Decl., ¶ 6; Mullis Dep. at 75.</p>

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in the sample.	
17. By contrast, non-specific primers and enzymes will amplify <i>any</i> and <i>all</i> sequences present in the sample.	No dispute.
18. The random primers will bind to all of the sequences in the sample and non-specific replication enzymes will catalyze DNA synthesis at points throughout the entire lengths of the nucleic acid molecules present without regard to sequence.	No dispute.
19. In its TMA method, Gen-Probe uses two amplification enzymes that depend upon the presence of specific primers.	No dispute.
20. One of these enzymes is reverse transcriptase ("RT").	No dispute.
21. RT is a DNA polymerase that produces a complementary DNA strand copy of a single-stranded RNA or DNA that has a bound primer.	No dispute.
22. In TMA, RT produces complementary DNA from the target nucleic acids (or their	No dispute.

<p>1 complementary strands) only if the sequence-  2 specific primers first bind to a single strand of  3 RNA or DNA.  4</p>	
<p>5 23. If the target organism is not present in the  6 sample, the primers will be unable to bind to  7 the captured sequence and the RT will not  8 initiate synthesis.  9</p>	<p>Disputed. All nucleic acid amplification  techniques have some degree of nonspecificity.  See Persing Decl., ¶ 6; Mullis Dep. at 75.</p>
<p>10 24. Another specific primer used in Gen-  11 Probe's method also includes a specific  12 "promoter" sequence that is recognized by  13 another enzyme ("T7 RNA polymerase") that  14 binds specifically to that promoter sequence  15 to produce many RNA copies by  16 transcription.  17  18</p>	<p>No dispute.</p>
<p>19 25. A function "T7 promoter" is formed in  20 the course of the TMA process if, and only if,  21 (1) the primer finds and binds to its  22 complementary target sequence in the  23 captured target molecule so that the target  24 sequence is copied by reverse transcriptase  25 and (2) the second primer binds to the newly  26 synthesized DNA and DNA polymerase  27  28</p>	<p>Disputed. All nucleic acid amplification  techniques have some degree of nonspecificity.  See Persing Decl., ¶ 6; Mullis Dep. at 75.</p>

<p>1 makes the complementary DNA strand.</p> <p>2</p>	
<p>3 26. If this double-stranded, and hence</p> <p>4 functional, T7 promoter <i>is</i> formed as a result</p> <p>5 of these <i>two</i> primer binding and extension</p> <p>6 processes, then the T7 RNA polymerase used</p> <p>7 in Gen-Probe's HIV/HCV test will amplify</p> <p>8 the sequence attached to the T7 promoter</p> <p>9 sequence.</p> <p>10</p>	<p>No dispute.</p>
<p>11 27. The T7 RNA polymerase does not</p> <p>12 amplify other sequences present in the sample</p> <p>13 because they are not attached to a T7</p> <p>14 promoter sequence.</p> <p>15</p> <p>16</p>	<p>Disputed. All nucleic acid amplification</p> <p>techniques have some degree of nonspecificity.</p> <p>See Persing Decl., ¶ 6; Mullis Dep. at 75.</p>
<p>17 28. Thus, in Gen-Probe's HIV/HCV test, the</p> <p>18 T7 polymerase enzyme <i>specifically</i></p> <p>19 recognizes the T7 promoter sequence, which</p> <p>20 has been <i>specifically</i> attached to the target</p> <p>21 sequence by the binding of <i>specific</i> primers,</p> <p>22 and the T7 polymerase <i>specifically</i> amplifies</p> <p>23 only that sequence.</p> <p>24</p> <p>25</p>	<p>Disputed. All nucleic acid amplification</p> <p>techniques have some degree of nonspecificity.</p> <p>See Persing Decl., ¶ 6; Mullis Dep. at 75.</p>
<p>26 29. The process repeats in a cyclic fashion,</p> <p>27 only amplifying the particular target sequence</p> <p>28</p>	<p>Disputed. All nucleic acid amplification</p> <p>techniques have some degree of nonspecificity.</p>



1	of interest.	See Persing Decl., ¶ 6; Mullis Dep. at 75.
2		
3	30. Gen-Probe's amplification method	Disputed. All nucleic acid amplification
4	therefore safeguards against amplification of	techniques have some degree of nonspecificity.
5	non-target sequences and thus protects against	See Persing Decl., ¶ 6; Mullis Dep. at 75.
6	false positive results.	
7		
8	31. TMA functions in way that is	Disputed. See Persing Decl., ¶¶ 9-16.
9	substantially different than the way in which	
10	non-specific amplification functions.	
11		
12	32. Specific amplification methods	Disputed. Specific amplification methods can
13	commonly achieve <i>exponential</i> amplification	achieve either linear or exponential
14	of the target sequence, as compared with	amplification, depending on the reaction
15	linear amplification.	conditions and the techniques employed. See
16		Mullis Dep. at 102-03
17		
18	33. Sustained, significant, exponential	Disputed. Specific amplification methods can
19	amplification is a hallmark of specific	achieve either linear or exponential
20	amplification methods.	amplification, depending on the reaction
21		conditions and the techniques employed. See
22		Mullis Dep. at 102-03.
23		
24		
25	34. In contrast, the non-specific amplification	No dispute.
26	methods of Examples 4 and 5 of the '338	
27	patent admittedly achieve only linear	
28		

<p>1 amplification, not exponential amplification.</p>	
<p>2</p> <p>3 35. The non-specific amplification methods</p> <p>4 of Examples 5 and 6 also cannot achieve</p> <p>5 exponential amplification. Because random</p> <p>6 primers bind at various places along the</p> <p>7 nucleic acids present in the sample, the</p> <p>8 products of amplification are fragmented.</p> <p>9</p>	<p>Disputed. Example 6 of the '338 patent</p> <p>discloses a technique for achieving exponential</p> <p>amplification of a target nucleic acid. ('338</p> <p>patent, col. 31, line 55 to col. 32, line 7.)</p>
<p>10</p> <p>11 36. If these products were then subjected to</p> <p>12 another round of non-specific amplification,</p> <p>13 the resulting products would be smaller still.</p> <p>14</p>	<p>Disputed.</p>
<p>15 37. Multiple rounds of non-specific</p> <p>16 amplification thus diminish rapidly in</p> <p>17 efficiency, whereas multiple rounds of</p> <p>18 specific amplification produce extraordinarily</p> <p>19 large amounts of full size product nucleic</p> <p>20 acids in very short periods of time.</p> <p>21</p>	<p>Disputed. All nucleic acid amplification</p> <p>techniques have some degree of nonspecificity.</p> <p>See Persing Decl., ¶ 6; Mullis Dep. at 75.</p>
<p>22 38. Non-specific amplification using random</p> <p>23 hexamer primers results in fragmented nucleic</p> <p>24 acids, each of which contains the random</p> <p>25 sequences present in the primers.</p> <p>26</p> <p>27</p> <p>28</p>	<p>No dispute.</p>

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<p>39. The resulting products are thus heterogeneous and have undefined composition.</p>	<p>Disputed.</p>
<p>40. Such nucleic acids are unsuitable for most of the purposes for which homogeneous, specifically amplified nucleic acids of known composition are employed.</p>	<p>Disputed. In the context of the claimed invention, non-specific amplification techniques can amplify target nucleic acids in a manner sufficient to permit their detection as part of a diagnostic assay.</p>
<p>41. As a result, Gen-Probe's TMA method also does not yield the same result as that obtained with non-specific amplification.</p>	<p>Disputed. See Persing Decl., ¶¶ 9-16.</p>
<p>42. The Court has previously noted that the specification of the '338 patent contains no reference to any specific amplification techniques. To the contrary, the specification clearly suggests that the claimed amplification techniques of the invention don't require the use of specific primers necessary for specific amplification.</p>	<p>Vysis disputes the implication that specific amplification techniques are excluded from the claims of the '338 patent.</p>

1 43. This absence in the '338 patent of any  
2 disclosure of specific amplification techniques  
3 was not accidental or unintended. To the  
4 contrary, Gene-Trak Systems, Vysis'  
5 predecessor-in-interest, and its employed  
6 inventors were well aware of the specific  
7 amplification techniques such as PCR. In  
8 fact, the admitted focus of the inventors'  
9 effort leading to the disclosure in the '338  
10 patent was to find something "different" from  
11 specific amplification. For example, inventor  
12 Jon Lawrie testified that the patent was meant  
13 to cover new amplification methods using  
14 non-specific primers, not already-known  
15 methods such as PCR:

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19 Q. Can you recall any reason that a  
20 reference to PCR might have been  
21 intentionally omitted from the  
22 patent application?

23 A. Yes....

24  
25  
26 Q. If there's no reference in the  
27 ['338] patent to combining target  
28 capture with PCR, do you have any  
explanation as to why it is not there?

Vysis disputes there is an absence of any  
disclosure of specific amplification in the '338  
patent. Vysis does not dispute that Dr. Lawrie  
made the quoted statements in his deposition,  
but disputes the relevance of those statements  
to the determination of infringement under the  
doctrine of equivalents.

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A. I believe that it was a separate, the thought behind this [referring to the '338 patent] was coming up with new methods of amplification, not old ones.

Q. For the purposes of what you just said you classify PCR as an old method of amplification?

A. PCR itself was described in the patent, issued patent [e.g., it was an "old" method].

Q. And your understanding of the 338 patent was that it was directed to other methods of amplification?

A. The, it was, it was directed to the methods disclosed by, you know, the *methods separate from PCR*.

44.. Inventor King also stated the inventors' purpose and also distinguished non-specific amplification from PCR:

Q. From a high level perspective, what were the discussion topics

Vysis does not dispute that Dr. King made the quoted statements in his deposition, but disputes the relevance of those statements to the determination of infringement under the doctrine of equivalents.

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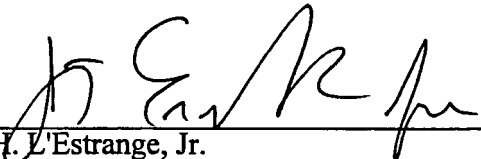
<p>addressed during this meeting?</p> <p>A. I think that at the highest level we were looking for amplification methods <i>that did not involve PCR amplification.</i></p> <p>(King Depo. At 45:10-15 (emphasis added).)</p> <p>Q. Okay. So the purpose -- the general purpose of the discussion as I understand it that took place at Gene-Trak among the four doctors was to identify -- in general identify an amplification technique that would amplify low concentrations of target nucleic acids in a sample, correct?</p> <p>A. Yes.</p> <p>Q. And as I understand your testimony, you wanted to find a technique <i>that was different from PCR</i>, correct?</p> <p>A. Yes.</p>	<p>doctrine of equivalents.</p>
<p>45. As this testimony suggests, PCR was well known to the inventors and the scientific</p>	<p>No dispute.</p>

<p>1 community at large. Dr. Kary Mullis invented  2 PCR in 1983, for which he received the Nobel  3 Prize in Chemistry. Dr. Mullis and his  4 colleagues publicly described PCR at a  5 scientific meeting in the summer of 1985 and  6 published their discovery in December 20,  7 1985.</p>	
<p>10 46. James Richards, Gene Trak's Director of  11 Business Development and Licensing, admits  12 that, within the scientific community, PCR  13 was immediately "big news."</p>	<p>No dispute.</p>
<p>15 47. One of the reasons that the '338 inventors  16 sought to find something "different" from  17 specific amplification techniques such as PCR  18 was due to Gene Trak's concern that it could  19 not obtain a license from Cetus Corp. to use  20 PCR. Cetus Corporation, which employed  21 Dr. Mullis, originally owned the rights to  22 PCR. Gene-Trak sought a license from Cetus,  23 but its requests were rejected.</p>	<p>No dispute.</p>
<p>26 48. The view of the fundamental difference  27 between non-specific and specific</p>	<p>Vysis disputes the statement that there is a  "fundamental difference between non-specific</p>

<p>1 amplification techniques was shared not only</p> <p>2 between the inventors but with Gene-Trak</p> <p>3 scientific management as well. In particular,</p> <p>4 in a letter he wrote in 1989, Dr. Richards,</p> <p>5 pointedly contrasted the '338 patent's method</p> <p>6 of non-specific amplification with other</p> <p>7 known specific methods that used specific</p> <p>8 primers or promoters:</p> <p>9</p> <p>10</p> <p>11 Cetus, Sibia/Salk, Biotechnica, etc.</p> <p>12 all claim specific primers for</p> <p>13 amplification <i>whereas the present</i></p> <p>14 <i>invention claims uses of the</i></p> <p>15 <i>opposite, namely, non-specific</i></p> <p>16 <i>primer or promoters....</i></p>	<p>and specific amplification techniques." See</p> <p>Persing Decl., ¶¶ 5 -16. Vysis also disputes</p> <p>that the independent claims of the '338 patent</p> <p>ever recited non-specific primers or promoters.</p>
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17 Date: November 8, 2001

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